

**Table S1: Yeast strains used in these analyses.**

*BY4741: MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0*  
*trf4 $\Delta$ : MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, trf4 $\Delta$ ::kanMX*  
*trf5: MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, trf5 $\Delta$ ::kanMX4*  
*trf4 $\Delta$ , GAL::trf5: MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, trf4 $\Delta$ ::kanMX4, HisMX6-pGAL-3HA::trf5*  
*rrp6 $\Delta$ : MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, rrp6 $\Delta$ ::natMX*  
*rrp6 $\Delta$ , trf4 $\Delta$ : MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, trf4 $\Delta$ ::kanMX, rrp6 $\Delta$ ::natMX*  
*rrp6 $\Delta$ , trf5: MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, trf5 $\Delta$ ::kanMX4, rrp6 $\Delta$ ::natMX*  
*GAL::mtr4: MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, HisMX6-pGAL-3HA::mtr4*  
*GAL::rrp41: MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, HisMX6-pGAL-3HA::rrp41*  
*GAL::rrp44: MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, HisMX6-pGAL-3HA::rrp44*  
*TRF5-TAP: MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, TRF5-TAP-URA3*  
*TRF5-TAP DADA: MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, TRF5 (D233A,D235A)-TAP-URA3*  
*TRF5-TAP trf4D: MAT<sub>a</sub>; his3 $\Delta$ 1; leu2 $\Delta$ 0; lys2 $\Delta$ 0; ura3 $\Delta$ 0, trf4 $\Delta$ ::kanMX, TRF5-TAP-URA3*

**Table S2. Hybridization probes used in this work. Major RNA species detected in Figs. 1-3 are indicated in brackets.**

003 (27S<sub>A2</sub>, 23S pre-rRNA) TGTTACCTCTGGGCCC  
004 (20S, 23S pre-rRNA) CGGTTTTAATTGTCCTA  
007 (25S) CTCCGCTTATTGATATGC  
008 (18S) CATGGCTTAATCTTTGAGAC  
015 (5.8S) TTTCGCTGCGTTCTTCATC  
041 (5S) CTA CTCTCGGTCAGGCTC  
020 (7S, 5.8S+30, 6S, 27S pre-rRNA) TGAGAAGGAAATGACGCT  
033 (5' ETS) CGCTGCTCACCAATGG  
202 (U14) TCACTCAGACATCCTAGG  
214 (U24) TCAGAGATCTTGGTGATAAT  
403 (PGK1) ACCGTTTGGTCTACCCAAGTGAGAAGCCAAGACA  
499 (TSA1) GGAGTATTCGGAGTCAGTGGAGGCGAAAAGAACT

**Table S3. Other oligonucleotides used in this work.**

RRP41F1 UP45: ATTTACAAAAAACTTTAGTGCCATAACTACAGCAGGATCATCATGAATT  
CGAGCTCGTTTAAAC  
RRP41R1 DN45: ATCGAGACGTAGCCCTTCTGGCGAGTATATTTCTAGTCTTGACATG  
CACTGAGCAGCGTAATCTG

RRP44F1 UP45: AACGAGTTTTATTTATCATACTTGCATCATACAGGCCAAAACAAC  
GAATTCGAGCTCGTTTAAAC

RRP44R1 DN45: ATCTGCAAGTCTCTTCCGTCTGGGGGCGATAGCGGGAACTGA  
CATGCACTGAGCAGCGTAATCTG

TRF5F2 UP45: GCTCAAACGAGAAGGGACTACTGGCTCTCTAAAGGCCAGG  
CTCTTTCCATGGAAAAGAGAAG

TRF5R2 DN45: TATTCTTGTATAAATAGTAAATAGTCTATAAGAGTCTATATTG  
TGTACGACTCACTATAGGG

TRF5 MUT1: TTGCCGGGTTCTGCAATTGCATGTGTCGTAAAC

TRF5 MUT2: GTTTACGACACATGCAATTGCAGAACCCGGCAA

## Legend to Supplementary Figure S1

A: Structure of the yeast pre-rRNA and locations of oligonucleotides used.

The 35S pre-rRNA contains the sequences of the mature 18S, 5.8S and 25S rRNAs, which are separated by internal transcribed spacers 1 and 2 (ITS1 and ITS2) and flanked by the 5' and 3' external transcribed spacers (5'ETS and 3'ETS).

B: The yeast pre-rRNA processing pathway. A complex processing pathway converts the 35S pre-rRNA primary transcript to the mature rRNAs. In wild-type cells, the 35S pre-rRNA is cleaved at site  $A_0$  producing the 33S pre-rRNA. This molecule is rapidly cleaved at site  $A_1$  to produce the 32S, which is cleaved at site  $A_2$  releasing the 20S and 27SA<sub>2</sub> pre-rRNAs. The 20S pre-rRNA is exported to the cytoplasm where it is cleaved at site D, by an unidentified enzyme, to generate the mature 18S rRNA. 27SA<sub>2</sub> is processed via two alternative pathways. It is either cut at site  $A_3$  to generate 27SA<sub>3</sub>, which is then trimmed to site B<sub>1S</sub>, producing 27SB<sub>S</sub>. Alternatively, it can be processed to 27SB<sub>L</sub> by an as yet unknown mechanism. 27SB<sub>S</sub> and 27SB<sub>L</sub> are matured to the 5.8S and 25S following identical pathways. Cleavage at site C<sub>2</sub> generates the 7S and 26S pre-rRNAs. The 7S pre-rRNA is digested 3' to 5' to 6S pre-rRNA and then to the mature 5.8S rRNA. The 26S pre-rRNA is digested 5' to 3' to the mature 25S rRNA. During this maturation the rRNA regions will assemble with the 80 ribosomal proteins, and the pre-rRNAs will transiently associate with ~170 protein processing and assembly factors and ~70 snoRNAs.

C: Cartoons of the predicted structures of aberrant pre-rRNA species detected in *rrp6Δ* strains. The 23S and 21S RNAs are aberrant intermediates, which are generated by premature cleavage at site  $A_3$ . We have not mapped the ends of the 17S' species but analyses of other ribosome synthesis mutants identified a similar RNA that has a 3' end at site  $A_3$  and 5' ends at heterogeneous positions within the mature 18S rRNA sequence. This may arise from inefficient 5' degradation of the 23S and 21S species.

